

FUTURISTIC BIOTECHNOLOGY

https://fbtjournal.com/index.php/fbt ISSN(E): 2959-0981, (P): 2959-0973 Vol 05 Issue 02, (April-June, 2025)



Original Article



Mathematical Modeling of DNA Yield Using Box-Behnken Design for Key Extraction **Parameters**

Saeeda Zia¹, Farwa Afzal¹, and Rashid Saif^{2*}

Department of Sciences and Humanities, National University of Computer and Emerging Sciences, Lahore, Pakistan

ARTICLE INFO

Keywords:

DNA Yield Modeling, Response Surface Design, Mathematical Biology, Biomathematical Modeling

Zia, S., Afzal, F., & Saif, R. (2025). Mathematical Modeling of DNA Yield Using Box-Behnken Design for Key Extraction Parameters: Box-Behnken Design for Optimizing DNA Extraction Yield. Futuristic Biotechnology, 5(2), 56-61. https://doi.org/10.54393/ fbt.v5i2.174

*Corresponding Author:

Rashid Saif

Department of Biotechnology, Qarshi University, Lahore, Pakistan rashid.saif37@gmail.com

Received Date: 3rd April, 2025 Revised Date: 2nd June, 2025 Acceptance Date: 9th June, 2025 Published Date: 30th June, 2025

ABSTRACT

The current research was initiated to model an enzyme-assisted inorganic salt-based DNA extraction protocol to obtain maximum yield from peripheral whole blood samples. Objectives: DNA Key extraction parameters were considered for model optimization such as incubation temperature (54,56,58°C), time (8,10,12hrs.), and proteinase K enzyme concentrations (8,10,12µL). Methods: Box-Behnken Design (BBD) was employed to model and simulate the extraction process with three factors/variables and three levels each, which is one of the widely used functional approach of response surface methodology (RSM). With this selected design, RSM entails a mathematical model with experimental data to optimize DNA yield. Results: The best fit simulation settings yielded the maximum DNA at an incubation temperature of 56°C for 10hours with a proteinase K enzyme volume of 10µL. Utilizing these parameters, a yield of 300ng/µL was obtained. A confirmation run under these settings validated the prediction, with observed yields closely matching the model's estimates (p<0.05), indicating strong agreement between the model and experimental outcomes. Conclusions: Regression modeling identified the role of three key factors for optimal DNA yield. Additional extraction parameters can be integrated to develop a more robust and controlled model, ensuring efficient use of limited crime scene samples and reagents in forensic investigations.

INTRODUCTION

The DNA isolation of the specimens is a very important process in modern biomedical studies and diagnosis, and has wide-ranging applications that include genetic studies, as well as in identifying diseases. With the growing need to achieve efficiency and reliability in extraction methodologies, researchers are considering new methods of extraction using materials like magnetic nanoparticles and silica matrices as an overcoming of limitations inherent in traditional methods. This paper is concerned with the mathematical modeling of the response surface modeling (RSM) of Box-Behnken Design (BBD) of the entire whole-blood by inorganic salt-based DNA extraction, which is a basic traditional technique, to comprehend and optimize the parameters that can in turn enhance its

economic viability in terms of time, reagents and cost saving [1-3]. DNA extraction procedures often use organic solvents, including phenol-chloroform-isoamyl alcohol (PCI), which is renowned due to its efficiency in getting high yields of DNA, but is very expensive. On the contrary, inorganic salt-based techniques make use of cheaper substances such as sodium chloride (NaCl). The optimization of extraction methods based on inorganic is, however, still a research topic. Regression, ANOVA, and the R2 values are mathematical models that are critical in explaining the complex interplay between inorganic compounds and biological elements and offer a logical structure of forecasting and controlling the results. This can be a groundbreaking strategy of DNA extraction,

²Department of Biotechnology, Qarshi University, Lahore, Pakistan

especially in low-resource environments, to facilitate the study of genetic medicine, forensics, and molecular diagnostics research in the third world developing nations [2, 4]. The use of inorganic DNA extraction techniques has a wide range of reagents and protocol parameters, such as incubation temperature, time period, and the most important enzyme concentration or proteinase K, which is the most important in increasing the yield of DNA in whole blood samples. Other statistical methods, like RSM, are used in this study to perform the optimization of DNA extraction parameters rigorously. The key goal is to identify conditions that will maximize the yield of DNA with the use of BBD surface design. This research plays an important role in the establishment of the optimum values of the minimum required incubation temperature, duration of time, and concentration of proteinase K enzyme to maximize the yield of DNA before experimentation [5, 6]. This study aims to develop an integrated model that synthesizes principles from chemistry, biology, statistics, and mathematics to optimize key parameters.

METHODS

This study used Box-Behnken Design (BBD) to model and simulate the extraction process with three factors and three levels each. The study was conducted from August 2023 to July 2024 at Decode Genomics, Lahore, Pakistan and Ethical approval was obtained from the institutional review board (Ref. No. DG-adm-127). For our experiment, DNA was extracted from 200 µL of whole blood using a saltbased inorganic method. After lysis with TE buffer, SDS, and Proteinase K, 6M NaCl was added to pellet protein debris, incubating further on ice as well as centrifuging. DNA was precipitated in the supernatant using chilled isopropanol, which was monitored via gentle inversion. DNA pellets were washed sequentially three times with ethanol before a final harvest to remove impurities. Pure DNA was assayed using gel electrophoresis for intactness, as well as its concentration was determined [7, 8]. The following process variables and their ranges were used regarding the key DNA extraction reagents and chemicals

Table 1: Process variable labels and levels

Labels	Variables (units)	Levels			
	variables (units)	-1	0	1	
А	Incubation Temperature (° $\mathcal C$)	54	56	58	
В	Incubation Time (Hour)	8	10	12	
С	Proteinase-K Enzyme (μL)	8	10	12	

These variables include levels, variables (incubation temperature (°C), incubation time (hours), and proteinase K enzyme (µL). Each variable is categorized into three levels: -1, 0, and 1. For temperature, the range spans from 54°C, 56°C, 58°C. Time varies from 8, 10, and 12 hours, while proteinase K enzyme volume ranges from 8, 10 and 12 μ L. This table serves as a reference for understanding the experimental conditions and their corresponding parameter ranges, crucial for ensuring consistency and repeatability in the experiment's outcomes. BBD was used with Design Expert Software to design a DNA extraction optimization protocol. The software-designed experimental conditions that were simulated to run practical extraction trials as well as to simulate response surface to get a regression line (Table 2).

Table 2: Box-Behnken Experimental Design

Sr. No.	Α	В	С	Yield
1	56.00	10.00	10.00	350
2	56.00	8.00	8.00	90
3	56.00	10.00	10.00	350
4	54.00	10.00	8.00	110
5	56.00	10.00	10.00	350
6	58.00	10.00	12.00	310
7	56.00	8.00	12.00	270
8	56.00	10.00	10.00	350
9	56.00	10.00	10.00	350
10	54.00	10.00	12.0	100
11	58.00	10.00	8.00	180
12	58.00	12.00	10.00	300
13	56.00	12.00	8.00	150
14	54.00	12.00	10.00	220
15	54.00	8.00	10.00	200
16	56.00	12.00	12.00	400
17	58.00	8.00	10.00	180

Through systematic exploration of these combinations, we conducted 17 experiments aimed at DNA vield from wholeblood samples followed by DNA quantification. Response Surface Methodology (RSM) is a statistical technique used in modeling and multi-variable process optimization. In our research work, we applied the RSM to optimize the DNA extraction protocol, considering the interactive effects of DNA concentration, incubation time, and incubation temperature on the yield. The technique allows the establishment of predictive models to identify the optimum conditions for efficient and reproducible DNA recovery [9, 10]. The BBD embedded into the RSM was applied to optimize DNA extraction conditions by the exploration of the linear and the quadratic effect of three variables. Three levels per parameter (-1, 0, and +1)decrease the number of run experiments economically in comparison with full-factorial experiments. In combination with the RSM, it enabled the building of a predictive model for the identification of the condition providing the optimum DNA recovery [11]. The secondorder polynomial model used in response surface methodology (RSM) can be expressed as: $Y = \beta_0 + \sum_{j=1}^k \beta_j X_j + \sum_{j=1}^k \beta_{jj} X_j^2 + \sum_{i=0}^k \sum_{j=2}^k \beta_{ij} X_i X_j + e_j \, (1).$

After putting all the values in equation (1) and simplifying we get:

 $Y = \beta_0 + \beta_1 X_1 + \beta_2 X_2 + \beta_3 X_3 + \beta_{11} X_1^2 + 2\beta_{22} X_2^2 + 2\beta_{33} X_3^3 + \beta_{12} X_1 X_2 + \beta_{13} X_1 X_3 + 2\beta_{23} X_2 X_3 + e(2)$ Now, by substituting the actual experimental factors: $X_i = A$ (Incubation Temperature), $X_2 = B$ (Incubation Time), $X_3 = C$ (Proteinase K Volume) The model becomes:

 $Y = \beta_0 + \beta_1 A + \beta_2 B + \beta_3 C + \beta_{11} A^2 + 2\beta_{22} B^2 + 2\beta_{33} C^2 + \beta_{12} A B + \beta_{13} A C + 2\beta_{23} B C + e \ (3)$ Where Y is the predicted DNA yield $\frac{ng}{n\mu}$, β_0 is the intercept, and β_i , β_{ii} , β_{ij} are the linear, quadratic, and interaction coefficients, respectively and ε is the residual error term. A second order polynomial regression model was applied to correlate the response variable (DNA yield) and the independent variables: proteinase K enzyme concentration, time of incubation, and incubation temperature. The model applied in the Response Surface Methodology (RSM) was as follows: proteinase K enzyme concentration, time of incubation, and incubation temperature. The model applied in the Response Surface Methodology (RSM) was as follows: $Y = \beta_0 + \sum \beta_i X_i + \sum \beta_{ij} X_i^2 + \sum \beta_{ij} X_i X_i$ Analysis of variance (ANOVA) was performed to test the significance and adequacy of the model. The model performance was tested based on the values of R^2 , adjusted R², and predicted R² and also from lack-of-fit tests. All statistical analysis and model fitting were performed by Design Expert software (Stat-Ease Inc., USA). To authenticate the validity and predictive capacity of the constructed RSM model, a further experimental run was performed according to the optimal settings indicated by the model. The optimal levels of the concentration of the enzyme proteinase K, the time of incubation, and the temperature of incubation used for this purpose were derived from the numerical optimization function of the Design Expert software. The DNA extraction was performed according to the above-mentioned optimized parameters and the extracted DNA yield was validated against the corresponding value anticipated from the model to check the model's accuracy [12]. Model selection applied sequential sum of squares and model summary statistics. The quadratic model was the preferred model of fit because its p-value was significant, its R^2 and it's adjusted R² The values were reasonable, and it was able to capture linear and interaction effects between the variables that influenced DNA yield.

RESULTS

This equation represents the response function used for the estimation of DNA extraction yield from whole blood. The equation contains various independent variables, i.e., incubation temperature (A), incubation time (B), and Table 3: Sequential Model Sum of Squares

Proteinase K Enzyme Concentration (C), with corresponding coefficients estimated from the experimental data. The equation follows the following form: Y= 350.00 + 32.50A + 51.25B + 68.75C + 45.00AB $+35.00AC +17.50BC - 98.75A^2 - 46.25B^2 - 76.25C^2$. Here, (Y) represents the DNA extraction yield in a ng/uL. The independent variables and the interaction effects between the variables show the influence of the variables on the extraction. The linear and nonlinear relationships between the variables and the extraction yield represented by the quadratic terms (such as A2, B2, and C2) give insight about the potential nonlinear relationships. The above equation serves as the predictive model to predict DNA extraction yield from the specified experimental condition. The adequacy of the Box-Behnken model constructed was statistically validated, showing a very good fit with a high R² value of 0.975, an adjusted R² of 0.958, and a predicted R² of 0.922, while the lack-of-fit test value of 0.231 indicated an insignificant lack-of-fit, confirming that the model can be applied for the estimation of DNA yield under different extraction conditions. The relationship between incubation temperature, incubation time, and Proteinase K enzyme concentration and DNA yield was captured by the model, with linear, quadratic, and interaction terms indicating that the effects were not necessarily linear and allowing for the determination of optimal extraction conditions. Regression analysis showed that incubation temperature had a significant effect on DNA yield, with the response surface being concave and the optimal yield achieved at approximately 56°C; deviations to 54°C or 58°C resulted in decreased yield, reflecting the heat sensitivity of the enzymatic lysis reaction. Incubation time also significantly affected DNA yield, with a parabolic response surface and optimal yield at 10 hours, while shorter or longer durations reduced efficacy. Proteinase K volume exerted a significant influence, with the highest yield at 10 µL and lower or higher volumes leading to reduced yield due to incomplete lysis or over-digestion. Sequential sum of squares analysis and model summary statistics indicated that the quadratic model was the most appropriate for predicting DNA yield, as it was statistically significant, captured the relationships between variables effectively, and demonstrated a suitable fit with $R^2=0.8751$ and adjusted R²=0.7146, while linear, 2FI, and cubic models were less suitable due to lack of explanatory power, aliasing, or inconsistencies between predicted and observed responses (Table 3).

•	·					
Source	Sum of Squares DF Mean Square		F Value	Prob>F	Remarks	
Sequential Model-Sum of Squares for Total Yield						
Mean	1.028 x10 ⁰⁰⁶	1.00	1.028 x10 ⁰⁰⁶	_	_	_
Linear	67275.00	3.00	22425.00	2.43	0.1118	Suggested

2F1	14225.00	3.00	4741.67	0.45	0.7238	_		
Quadratic	82336.76	3.00	27445.59	8.22	0.0108	Suggested		
Cubic	23375.00	3.00	7791.67	6.366 x10007	<0.0001	Aliased		
Residual	0.000	4.00	0.000	_	-	-		
Total	1.21 x10 ⁰⁰⁶	17.00	71470.59	_	_	-		
	Model Summary Statistic							
Source	Std. Deviation	R ²	Adjusted R ²	Predicted R ²	PRESS	Remarks		
Linear	96.05	0.3594	0.2115	-0.0618	1.988x10 ⁰⁰⁵	-		
2F1	102.82	0.4353	0.0965	-0.7405	3.258x10 ⁰⁰⁵	_		
Quadratic	57.79	0.8751	0.7146	-0.9977	3.740x10 ⁰⁰⁵	Suggested		
Cubic	0.000	1.0000	1.00000	_	I	Aliased		

ANOVA outcomes indicated the statistically significant fitted model (p = 0.0180), so the variation in DNA yield could be well explained using the selected factors. Incubation time (B) and Proteinase K volume (C) indicated significant effects (p=0.0405 and p=0.0120), respectively, while temperature (A) indicated a moderate effect (p=0.1557). Of particular interest was the statistical significance of the second-order terms A² and C², indicating non-linear behavior of the parameters. The interaction terms (AB, AC, BC) indicated statistical non-significance. The model indicated a large coefficient of determination (R²=0.8751), showing significant agreement between the predictive and the observed responses. An adequate precision (AP=5.895) confirmed the signal-to-noise ratio of the model being satisfactory for the exploration of the design space (Table 4).

Table 4: ANOVA for Response

Source	Sum of Squares	DF	Mean Square	F- Value	p- Value		
Model	1.63x10 ⁰⁰⁵	9	18204.08	5.45	0.0180		
A-Temperature	8450.00	1	8450.00	2.53	0.1557		
B-Time	21012.50	1	21012.50	6.29	0.0405		
C-Proteinase k	37812.50	1	37812.50	11.32	0.0120		
AB	8100.00	1	8100.00	2.43	0.1633		
AC	4900.00	1	4900.00	1.47	0.2651		
BC	1225.00	1	1225.00	0.37	0.5638		
A^2	41059.21	1	41059.21	12.30	0.0099		
B ²	9006.58	1	9006.58	2.70	0.1445		
C ²	24480.26	1	24480.26	7.33	0.0303		
CV	23.50						
AP	5.895						
PRESS	3.740+005						
Mean	245.88						
R ²	0.8751						
Adj-R ²	0.7146						

The next graphs provide the combined diagnostic plots employed to check for the fitness of the fitted regression model. The normal probability plot of studentized residuals indicates that the residuals are normally distributed since they lie along a straight line. The predicted vs. actual plot shows a good correlation between observed and model-predicted values, with data points tightly scattered around

the diagonal line, indicating the accuracy of the model. The residuals vs. predicted plot shows a random distribution of residuals, indicating homoscedasticity and the lack of systematic error. Both plots confirm that the regression model is statistically valid and accurate for prediction. (A) Normal probability plot. Blue circles closely follow the reference line, indicating normally distributed residuals. (B) Predicted vs. actual values. Green triangles lie near the diagonal, showing strong agreement between observed and predicted DNA yield. (C) Residuals vs. predicted values – Red squares are randomly scattered around zero, confirming homoscedasticity and model validity. Axes represent expected vs. actual residuals (A), actual vs. predicted yield (B), and predicted yield vs. residuals (C) (Figure 1).

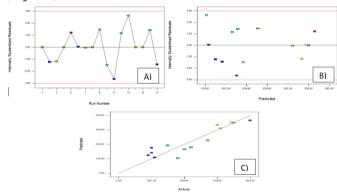


Figure 1: (A) Normal Probability Plot. (B) Predicted Vs. Actual Values. (C) Residuals Vs. Predicted. Axes Represent Expected Vs. Actual Residuals (A), Actual Vs. Predicted Yield (B), and Predicted Yield Vs. Residuals (C)

A three-dimensional scatter plot was created to demonstrate the interaction effects of incubation temperature, incubation time, and Proteinase K enzyme concentration on DNA yield using the Box-Behnken experimental design. The plot visually shows a nonlinear response surface with the highest DNA yield (~350–400 ng/µL) at points closest to the center values of 56°C , 10 hours, and 10 µL, validating the regression model's prediction. Deviation from these optimal conditions resulted in a drop in yield, validating the occurrence of quadratic effects and that there is a very small range within

which optimal extraction takes place. The visualization serves to reinforce that incubation time and enzyme concentration, specifically, need to be precisely balanced to preclude under- or over-digestion, both of which have a detrimental impact on yield. The even gradient of DNA concentration through the plotted surface gives a clear, intuitive view of the sensitivity of the extraction protocol to changes in each parameter. Generally, the plot reinforces the statistical results and verifies the robustness of the optimized conditions that were identified by the model (Figure 2).

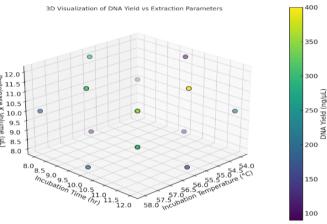


Figure 2: Three-Dimensional Scatter Plot

DISCUSSIONS

The optimization of the DNA extraction process by Box-Behnken design (BBD) showed that incubation temperature, incubation time, and Proteinase K enzyme concentration had a great nonlinear influence on the generation of the DNA, as had been previously reported [13]. The high-predictive quadratic regression model (R2=0.8751) illustrated that even minor alterations to the optimal conditions (56°C, 10 hours, 10 µL) had a major impact on the DNA recovery, which shows the sensitivity of enzymatic activity [14, 15]. The results are in line with the earlier optimization experiments with the use of factorial design and response surface methodology, which found the same quadratic behavior in the course of enzymatic extraction [16-18]. The concentration of proteinase K is critically important in determining extraction yield, due to the existence of two opposing conditions between total lysis and possible over-digestion [14, 16]. Comprehensively, the paper highlights the usefulness of response surface methodology (RSM) and Box-Behnken design in the effective determination of optimal conditions using a limited number of experimental programs to increase reproducibility and cost-efficiency [18, 19]. The improved salt-based extraction process was found to be appropriate in extracting high-quality DNA using a small amount of whole blood, and it is possible that it can be used in clinical diagnostics, forensic cases, and genetic studies [7, 20].

Even though the model system was chicken blood, the principles realized in this case can be applied in other species after validation. The model can be further improved in future research by adding some parameters to it, like buffer composition, storage conditions, and sample type.

CONCLUSION

An enzyme-based DNA extraction of whole blood was optimized by box-Behnken design and response surface methodology. The quadratic equation (R 2=0.8751) established optimal conditions (56 0 C, 10 hours, 10 0 -1) to give maximum yield. Reliability was also experimentally validated, and provides a low-cost, fast, and repeatable alternative to commercial kits with low sample volumes.

Authors Contribution

Conceptualization: SZ, RS Methodology: SZ, FA, RS Formal analysis: SZ, FA, RS

Writing review and editing: SZ, FA, RS

All authors have read and agreed to the published version of the manuscript.

Conflicts of Interest

All the authors declare no conflict of interest.

Source of Funding

The authors received no financial support for the research, authorship and/or publication of this article.

REFERENCES

- Dwivedi S, Purohit P, Misra R, Pareek P, Goel A, Khattri S, et al. Diseases and Molecular Diagnostics: A Step Closer to Precision Medicine. Indian Journal of Clinical Biochemistry. 2017 Oct; 32(4): 374-398. doi: 10.1007/s12291-017-0688-8.
- Neal CJ, Zbinden ZD, Douglas ME, Douglas MR. [2] Reducing DNA Extraction Costs Through Factorial Design for the DNAdvance Kit. BMC Research Notes. 2024 Dec; 17(1): 397. doi: 10.1186/s13104-024-07063-
- [3] Zhang S, Wang Y, Liu T, Li X, Wang M, Qiu Y. Optimization of the Extraction Process of Effective Components of Eleutherococcus senticosus Using Mathematical Models. Journal of Food Quality. 2024; 2024: 5538373. doi: 10.1155/2024/5538373.
- [4] Crooke PS and Parl FF. A Mathematical Model for DNA Damage and Repair. Journal of Nucleic Acids. 2010; 2010: 352603. doi: 10.4061/2010/352603.
- Silva E, Rogez H, Larondelle Y. Optimization of Extraction of Phenolics from Inga edulis Leaves Using Response Surface Methodology. Separation and Purification Technology. 2007 Mar; 55(3):

- 381-387. doi: 10.1016/j.seppur.2007.01.008.
- [6] Gulas-Wroblewski BE, Kairis RB, Gorchakov R, Wheless A, Murray KO, et al. Optimization of DNA Extraction from Field-Collected Mammalian Whole Blood on Filter Paper for Trypanosoma cruzi (Chagas Disease) Detection. Pathogens. 2021 Aug; 10(8): 1040. doi: 10.3390/pathogens10081040.
- Tang ZF and McMillen DR. An Inhibitor-Free, Versatile, Fast, and Cheap Precipitation-Based DNA Purification Method. PLOS ONE. 2025 Apr; 20(4): e0317658.doi:10.1371/journal.pone.0317658.
- [8] Lynton-Jenkins JG, Chaine AS, Russell AF, Bonneaud C. Parasite Detection and Ouantification in Avian Blood Is Dependent on Storage Medium and Duration. Ecology and Evolution. 2023 Feb; 13(2): e9819. doi: 10.1002/ece3.9819.
- [9] Khuri Al and Mukhopadhyay S. Response Surface Methodology. WIREs Computational Statistics. 2010 Mar; 2(2): 128-149. doi: 10.1002/wics.73.
- [10] Djimtoingar SS, Derkyi NS, Kuranchie FA, Yankyera JK, et al. A Review of Response Surface Methodology for Biogas Process Optimization. Cogent Engineering. 2022 Dec; 9(1): 2115283. doi: 10.1080/ 23311916.2022.2115283.
- [11] Gerzsenyi TB, Ilosvai ÁM, Kristály F, Daróczi L, Owen MC, Viskolcz B, et al. Investigation and Optimization of DNA Isolation Efficiency Using Ferrite-Based Magnetic Nanoparticles. Biotechnology Reports. 2025 Mar; 37: e00904. doi: 10.1016/j.btre.2025.e00
- [12] Hussain MA, Ali A, Alsahli TG, Khan N, Sharif A, Haseeb MT, et al. Polysaccharide-Based Hydrogel from Seeds of Artemisia vulgaris: Extraction Optimization by Box-Behnken Design, pH-Responsiveness, and Sustained Drug Release. Gels. 2023 Jul; 9(7): 525. doi: 10.3390/gels9070525.
- [13] Tan R, Zhou S, Sun M, Liu Y, Ni X, He J, Guo G, Liu K, et al. Modeling and Optimization of Culture Media for Recombinant Helicobacter pylori Vaccine Antigen HpaA. Frontiers in Bioengineering and Biotechnology. 2024 Dec; 12: 1499940. doi: 10.3389/ fbioe.2024.1499940.
- [14] Martínez-de la Puente J, Ruiz S, Soriguer R, Figuerola J, et al. Effect of Blood Meal Digestion and DNA Extraction Protocol on the Success of Blood Meal Source Determination in the Malaria Vector Anopheles atroparvus. Malaria Journal. 2013 Dec; 12(1): 109. doi: 10.1186/1475-2875-12-109.
- [15] Afzal F, Zia S, Saif R. Mathematical Modeling Using Box-Behnken Response Surface Design for Optimal DNA Yield with Variable Parameters of Time, Temperature, and Proteinase K Concentration.

- bioRxiv. 2024 Jul; 2024-07. doi: 10.1101/2024.07.21. 604461.
- [16] Shahriar M, Hague MR, Kabir S, Dewan I, Bhuyian MA, et al. Effect of Proteinase K on Genomic DNA Extraction from Gram-Positive Strains. Stamford Journal of Pharmaceutical Sciences. 2011; 4(1): 53-57. doi: 10.3329/sjps.v4i1.8867.
- [17] Van Biesen N, Cools P, Meyers E. Comparison and Optimization of DNA Extraction Methods for Human DNA from Dried Blood Spot Samples, Pediatric Reports. 2025 Mar; 17(2): 30. doi: 10.3390/pediatric17 020030.
- [18] Goemaere I, Puni D, Harizai A, Woolston J, Thys S, Sterck K, et al. Response Surface Methodology to Efficiently Optimize Intracellular Delivery by Photoporation. International Journal of Molecular Sciences. 2023 Feb; 24(4): 3147. doi: 10.3390/ijms24 043147.
- [19] Yang JS, Dias FF, De Moura Bell JM. Optimizing Alkaline and Enzymatic Extraction of Black Bean Proteins: A Comparative Study of Kinetics, Functionality, and Nutritional Properties. Sustainable Food Technology. 2025; 3(1): 188-203. doi: 10.1039/D4FB00163J.
- [20] Verma A, Manchel A, Melunis J, Hengstler JG, Vadigepalli R. From Seeing to Simulating: A Survey of Imaging Techniques and Spatially Resolved Data for Developing Multiscale Computational Models of Liver Regeneration. Frontiers in Systems Biology. 2022; 2: 917191. doi: 10.3389/fsysb.2022.917191.